INTRODUCTION

Oral absorption of drugs is an extremely complicated process that is influenced by physiochemical properties of the drug formulation and physiological conditions of the gastrointestinal (GI) tract (1). Edwards in 1951 first proposed that drug dissolution would play a critical role in oral absorption, and this relationship has been of great interest in the past decades (2, 3). In vitro dissolution testing provides considerable information for in vivo drug performance by establishing an in vitro-in vivo relationship (IVIVR). Noticeably, when dissolution tests have inaccurate discrimination for the performance of candidate formulations between in vitro and in vivo studies, the predictive results may be completely misleading. Thus, it is desirable that changes in in vivo drug dissolution are reflected by the corresponding in vitro drug release test.

The oral absorption of Biopharmaceutics Classification System (BCS) class II drugs is mainly limited by solubility and dissolution, which may be required to establish the IVIVR (4, 5). In particular, weakly basic drugs are significantly influenced by physiological conditions in the GI tract, especially for dynamic pH conditions. Weak bases freely dissolve at gastric pH, but not at intestinal pH due to a supersaturation-precipitation process that greatly affects drug bioavailability. Some dissolution tests may employ simple and single-pH non-physiologic buffers that do not accurately reflect in vivo dynamic situations (6, 7). To bridge the gap between in vitro and in vivo conditions, many attempts have been made to replicate the GI process of weak bases by developing various biorelevant dissolution methods (8, 9). Owing to the lack of drug removal from the system and challenges associated with simulated absorption across the intestinal membrane, in vitro dissolution may overestimate the in vivo situation.

A biphasic dissolution test offers the advantage of maintaining sink conditions and having an absorptive phase, which has had increasing attention (10). A biphasic dissolution test can simulate drug dissolution and absorption in the GI tract by the implementation of an immiscible organic phase acting as an absorptive sink over the aqueous solution. Moreover, the setup and handling of biphasic tests are relatively simple and cost

ABSTRACT

The aim of this study was to develop a method for set up and optimization of a pH-gradient biphasic dissolution model by orthogonal test design in light of the correlation with published in vivo data of ketoconazole (KTZ). A pH-gradient biphasic dissolution test was designed with a sequential pH-gradient in the aqueous phase to simulate stomach, duodenum, jejunum, and ileum, and the organic phase was added in simulated small intestine conditions. The model was optimized by orthogonal test design with three factors and three levels and correlating with the published pharmacokinetic data of pure drug. The optimized dissolution conditions were 30 rpm, 100 mL of an organic volume, and pH 5.5, 6.5, and 6.8 in the pH-gradient aqueous phase in USP apparatus 2. Under these conditions, KTZ dissolution displayed a good linear relationship with in vivo absorption ($R^2 = 0.85$). This study indicates that this methodology is feasible to develop an in vivo predictive dissolution test.

KEYWORDS: pH-gradient biphasic dissolution test, in vitro-in vivo relationship (IVIVR), weakly basic drug, predictive dissolution test, ketoconazole, dissolution
effective, and drug release and partitioning are detected simultaneously within a single vessel (11). Previous studies have reported the development of various biphasic dissolution models and their correlation to pharmacokinetic (PK) data for different dosage forms (12–17). A biphasic system with a single pH of aqueous phase was developed by combining United States Pharmacopeia (USP) apparatus 2 and 4 methods to discriminate three celecoxib formulations and obtain a consistent ranking between drug concentrations of organic phase and in vivo PK parameters (12). To better mimic drug behaviors in the GI tract, a pH-adjusted biphasic dissolution system was developed to differentiate between four modified release formulations prepared with two weakly basic drugs (dipyridamole and BIMT 17) (13). The pH-adjusted biphasic test provided a ranking prediction with respect to in vivo absorption, but not in dissolution tests with different media at a constant pH. However, there are still some questions to be considered and solved. Firstly, there is a lack of studies that develop a method for setup and optimization of a biphasic test because the experimental parameters may have a critical influence on drug performance (11, 18). Secondly, pH-adjusted biphasic dissolution tests involve the addition of organic phase at gastric pH, which might result in an overestimated prediction of in vivo performance due to supersaturation of weak bases, which could lead to overestimation of oral bioavailability if the organic phase is added prematurely.

Based on the aforementioned concerns, ketoconazole (KTZ), a BCS II weakly basic drug with broad-spectrum antifungal activity, was selected as a model drug. The aim of the present study was to develop a method to set up and optimize a pH-gradient biphasic dissolution model by orthogonal test design in light of the correlation with published in vivo data of KTZ.

MATERIALS AND METHODS

Materials
KTZ was purchased from Wuhan Dahua Weiye Medicine Chemical Co., Ltd. (Wuhan, China). 1-octanol, hydrochloric acid (HCl), sodium dihydrogen phosphate dihydrate, sodium hydroxide (NaOH), and sodium chloride were obtained from Sichuan Kelun Pharmaceutical Co., Ltd. (Chengdu, China). Hard gelatin capsules (size 0) were donated by Suzhou Capsugel Ltd. (Suzhou, China). All other reagents used were of analytical grade.

Solubility Determination
Excessive KTZ was added to 10 mL of five different dissolution media (gastric buffer pH 2.0, phosphate buffer pH 5.5, 6.5, and 6.8, and 1-octanol). The solubility test was performed at 37 °C and 160 rpm for 24 h using a horizontal shaker (THZ-98AB, Shanghai Yiheng Scientific Instrument Co. Ltd, Shanghai, China) (n = 3). Saturated solutions were filtered through a 0.45-µm membrane filter. Drug concentration was measured by ultraviolet (UV) spectrophotometry at 224 nm (T6, Beijing Puxi General Instrument Co. Ltd, Beijing, China).

pH-Gradient Biphasic Dissolution Test
Considering the physiological pH-change in the GI tract which significantly influences KTZ dissolution, a pH-gradient biphasic dissolution test was proposed (Fig. 1). The test was performed in USP apparatus 2 (RCZ-6B3, Shanghai Huanghai Pharmaceutical Inspection Instrument Co. Ltd, Shanghai, China) combining with a pH controlling device (PHS-2F, Shanghai INESA Scientific Instrument Co., Ltd, Shanghai, China).

The gastric buffer was prepared with 250 mL of 0.01 N HCl with 0.1 M NaCl (pH 2.0) and used to simulate drug dissolution in the stomach for 30 min (19). The pH of the aqueous medium was adjusted to duodenal pH by using 5 M NaOH with a fresh sampling needle, and 1-octonal as the upper organic phase was added to simulate the intestinal membrane as an absorptive sink. 1-octonal was saturated with water prior to use to keep a constant volume during the experiment. Subsequently, the pH of the aqueous phase was readjusted to 6.5 to mimic the jejunum for 2 h, and then the final pH increased to 6.8 for 1 h. The temperature was maintained at 37 °C.

For the dissolution test, 100 mg of pure KTZ in a gelatin capsule was used with a sinker (n = 3); 3-mL samples were withdrawn manually from the aqueous phases at 5, 10, 20, 30, 40, 50, 60, 70, 80, 90, 105, 120, 150, 180, 210, and 240 min, while 2-mL samples were collected.
RESULTS AND DISCUSSION

Solubility

KTZ showed highly pH-dependent solubility in the physiologically relevant pH range (Table 2). As a weakly basic drug, KTZ had the highest solubility in the acidic medium, with 8.2 mg/mL dissolved in pH 2.0, which was used to mimic pH conditions of a healthy fasted stomach. In contrast, KTZ showed a dramatic decrease in solubility to 9.5 µg/mL in pH 5.5 buffer, following a slow fall from 3.9 to 3.5 µg/mL between pH 6.5 and pH 6.8. KTZ had high solubility of 5.6 mg/mL in 1-octanol due to its hydrophobic structure (logP 3.9) (23).

Table 2. Solubility of Ketoconazole in Dissolution Media at 37 °C

<table>
<thead>
<tr>
<th>Media</th>
<th>Solubility, mean ± SD (n = 3)</th>
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<tbody>
<tr>
<td>1-Octanol</td>
<td>5.6 ± 0.5 mg/mL</td>
</tr>
<tr>
<td>Gastric buffer pH 2.0</td>
<td>8.2 ± 0.2 mg/mL</td>
</tr>
<tr>
<td>PBS pH 5.5</td>
<td>9.5 ± 0.5 mg/mL</td>
</tr>
<tr>
<td>PBS pH 6.5</td>
<td>3.9 ± 0.6 µg/mL</td>
</tr>
<tr>
<td>PBS pH 6.8</td>
<td>3.5 ± 0.6 µg/mL</td>
</tr>
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PBS: phosphate buffer solution.

Optimization Study

The orthogonal experimental design was used to select the optimum dissolution conditions. The results of nine groups of experiments under different conditions are shown in Table 3. The k (k₁, k₂, or k₃) value was the mean of the sum of the three R² values for levels 1–3 of each factor, which revealed the change of predictive efficiency. The effects of levels 1–3 on drug dissolution were factor A (rotating speed of paddle): 1 > 2 > 3; factor B (organic volume): 2 > 1 > 3; factor C (pH gradients of aqueous phase): 1 > 3 > 2. The range (R) was the difference between the maximum and minimum value of k₁, k₂, and k₃ in the same table column, which reflected the impact degree of different factors. The larger the R value of one factor, the greater the impact on drug dissolution and in vivo prediction. According to the R value in Table 3, the order of three factors influencing drug dissolution and in vivo prediction was C > B > A. Therefore, the optimized dissolution parameters were A₁B₂C₁; namely, rotating speed is 30 rpm, the organic volume is 100 mL, and the pH-gradient aqueous phase is pH 5.5, 6.5, and 6.8.

ph-Gradient Biphasic Dissolution Test

To validate the prediction potential of the optimized dissolution test, KTZ dissolution was performed in the optimum pH-gradient biphasic dissolution test in triplicate. The dissolution profiles of pure drug are presented in Figure 2. Drug dissolution reached 100%
in the gastric buffer pH 2.0 due to high solubility in the acidic medium. After pH adjustment, drug concentration dramatically declined with increasing pH from 5.5 to 6.8 in the aqueous phase, even to below 20% in pH 6.8 medium (Fig. 2A). This could be attributed to rapid drug precipitation caused by low solubility of KTZ in the high pH media. In contrast, the amount of drug released gradually increased in the organic phase (Fig. 2B). It is well known that the small intestine is a main absorption site for most drugs (24). The addition of the organic phase after pH change was to simulate the intestinal absorption by removing dissolved drug from the aqueous phase and into the organic phase. Moreover, a good linear relationship was found between in vitro dissolution and in vivo $F_a$ ($R^2 = 0.85$) (Fig. 3), which was higher than the $R^2$ values obtained for all other conditions shown in Table 3.

**CONCLUSION**

An orthogonal design using experimental data combined with reported in vivo data was used to set up and optimize a pH-gradient biphasic dissolution test. The optimized experimental parameters were obtained based on $R^2$ of a linear relationship between in vitro and in vivo dissolution of KTZ. The highest $R^2$ was found when KTZ was tested in the optimized pH-gradient biphasic dissolution test, indicating feasibility of the method. In the next study,
different KTZ formulations will be used to further identify the discriminative and predictive ability of the proposed biphasic dissolution test.

![Graph showing relationship between in vivo dissolution of ketoconazole and in vivo absorption (Fa) calculated according to data reported in female rats (21).](image)

**Figure 3.** Relationship of in vitro dissolution of ketoconazole in the organic phase of optimized pH-gradient biphasic test plotted against in vivo absorption (Fa) calculated according to data reported in female rats (21).

**FUNDING**

This work was financially supported for purchasing materials and assay determination by the Science and Technology Research Program of Chongqing Municipal Education Commission (Grant No. KJQN201800820); the Chongqing Research Program of Basic Research and Frontier Technology (Grant No. cstc2019cyj-msxmX0753); the Scientific Research Project of Chongqing Technology and Business University (Grant No. 1951066); the open fund of the Key Laboratory of Natural Medicine Research of Chongqing Education Commission (Grant No. KFJJ2019093); the Open Project of Central Nervous System Drug Key Laboratory of Sichuan Province (Grant No. 200024-01SZ); and the Open Project of Central Nervous System Drug Key Laboratory of Sichuan Province (Grant No. KFJJ2019093); the Open Project of Central Nervous System Drug Key Laboratory of Sichuan Province (Grant No. 200024-01SZ).

**CONFLICT OF INTERESTS**

The authors disclosed no conflicts of interest related to this article.

**REFERENCES**


