

New Innovations in Testing Sustained-Release Tablets Using an Automated Dissolution System with Online Dilution

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ABSTRACT

This paper describes the development and validation of an automated method for the dissolution testing of metformin hydrochloride for use in the quality assurance program. Automation could provide significant improvements in the consistency of results compared with existing manual methods. Care and consideration were given to the automation of preparation of samples and standards in line with current manual procedures. Reproducibility and sample preparation accuracy were quantified, and results obtained were compared against tests performed manually.

The system was validated in accordance with the Chinese (1), European (2), and United States pharmacopeias (3) using salicylic acid. Salicylic acid testing for bath calibration also required dilution, which provided a direct comparison to the automated system used to test metformin hydrochloride tablets.

KEYWORDS: Dissolution; automation; online; validation; method development.

INTRODUCTION

Metformin hydrochloride is an oral antidiabetic drug used in the treatment of type 2 diabetes. It is also used in the treatment of polycystic ovary syndrome and has been investigated for other diseases where insulin resistance may be an important factor. Metformin is the only antidiabetic drug that has been conclusively shown to prevent the cardiovascular complications of diabetes (4, 5). In China, metformin hydrochloride oral solid dosages are manufactured by nearly 200 companies. The current dissolution methodology for testing metformin hydrochloride oral solid dosage is to collect samples, replace the sampling volume removed with fresh medium, dilute the sample, and measure by UV spectrophotometry.

In China, the National Institute for Food and Drug Control (NIFDC) is responsible for the regulation and quality of drugs. This includes post-market surveillance sampling and specification evaluation of imported drugs as well as drugs that are produced in China. We now report on a novel method for dissolution testing of solid dose metformin hydrochloride using an automated technique that we believe has advantages over methodology currently used by the NIFDC.

SYSTEM DESCRIPTION

A schematic representation of the new method and system configuration is shown in Figure 1. It consists of the ADT8 dissolution bath from Automated Lab Systems (Berkshire, UK), an ALS 8 Syringe Pump (used for sampling), ALS 3-port Sample Loss Replacement and Sampling Valves,

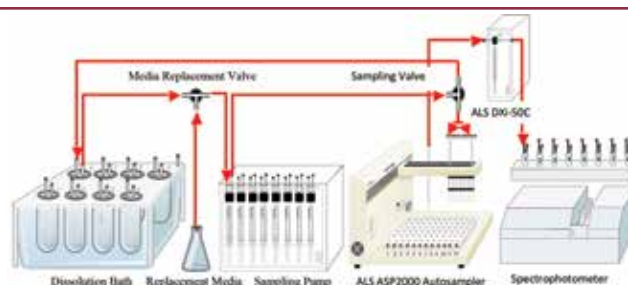


Figure 1. Schematic of the ASP2000 automated system connected directly to the UV spectrophotometer.

ALS ASP2000 Dual Probe XYZ Robotic Sampler, ALS DXi-50 Syringe Diluter, and PGI T70 Spectrophotometer (PG Instruments, Ltd.). Samples were filtered using in-line disc filters (Gelman Acrodisc GF 0.45- μ m) configured outside the dissolution vessels with Luer fittings for easy removal and replacement. Metformin hydrochloride exhibits UV absorption linearity up to about 15 mg/L at about 233 nm (7); therefore, 500-mg tablets require approximately 100-fold dilution, which is in line with current methodology (6).

The challenge for this test is diluting 1 in 100. The diluter is capable of single dilutions of 1 in 30 within 1% accuracy. Therefore, serial dilution was performed to achieve the higher dilution of 1 in 100. The ASP2000 XYZ Robotic Sampler fitted with two probes was used. Racks required for the analysis with serial dilution were configured using IDIS and ASP2000 teachable rack configuration, which allowed optimized positioning of samples to suit our requirements. Up to 44 time points, five stock standards, eight prepared standards, and two blanks, 100 mL each, can be configured for the ASP2000 Dual Probe XYZ Robotic Sampler. This

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virtual rack design allowed us to configure racks to realize the requirements to perform serial dilution. Three racks each accommodating 11 time points (11 × 7), 100-mL blank, and eight standards were configured (Figure 2).

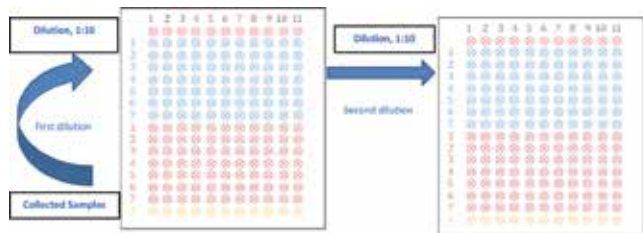


Figure 2. Schematic of racks configured to meet requirements for serial dilution.

The design of two probes realizes simultaneous collection and dilution using three-dimensional XYZ movement to access sample tubes anywhere on the ASP2000 Dual Probe XYZ Robotic Sampler. The left probe, with eight needles, was used to collect all samples simultaneously, while the right probe, with a single needle linked to the diluter, was used to dilute and inject the samples into the spectrophotometer.

The commercially available system is used by other companies for streamlining collection, dilution, and measurement for UV or HPLC automation. The schematic in Figure 3 shows the operation workflow. Prior to sampling, a selected volume of sample is pumped in circulation to the bath to equilibrate fresh sample solution in the lines. At the sampling time interval, the valve switches to the ASP2000 Multiple Probe, which moves to sampling tubes to dispense the desired volume of sample solution. The complete system is controlled by the IDIS software supplied by ALS. This software controls instruments from different manufacturers (baths from Agilent, Distek, Erweka, Pharmatest, Sotax; spectrophotometers from Agilent, Perkin Elmer, Jena, Shimadzu, Thermo; peristaltic and syringe pumps) and allows the user to configure methodologies using instruments from different sources for closed-loop UV analysis, collection, and HPLC analysis linked to ChemStation or Empower as well as to collect, prepare, and measure samples on UV followed by result calculations and reporting. Any combination of bath, pump, and spectrophotometer can be selected. Methodology is created by adding symbols representing the driver for the instruments onto the method desktop; linking the symbols by making connection to points on the symbols; and finally setting sampling times, temperature, and stirrer speed of the bath. During the analysis, raw data (absorbance in this case) acquired from the spectrophotometer, as well as temperature and stirrer speed from the bath, are stored to IDIS relational database immediately after collection.

The IDIS software optimizes the timings to make sure the time to the next sampling interval is not compromised. If there is enough time before the next samples are due, the ASP2000 single probe connected to the diluter is positioned at the appropriate tube and sample is aspirated,

then the ASP2000 moves to another tube and the diluter dispenses the sample with diluent to perform dilution. If there is not enough time to perform dilution or injection and measurement, the system defers this sequence until after the next collection, a process referred to as Deferred Data Collection by ALS. In practice, this deferment can occur indefinitely, especially for short time intervals, in which case the processing (dilution, injection, and measurement) will be performed at the end of the dissolution analysis time (Figure 3).

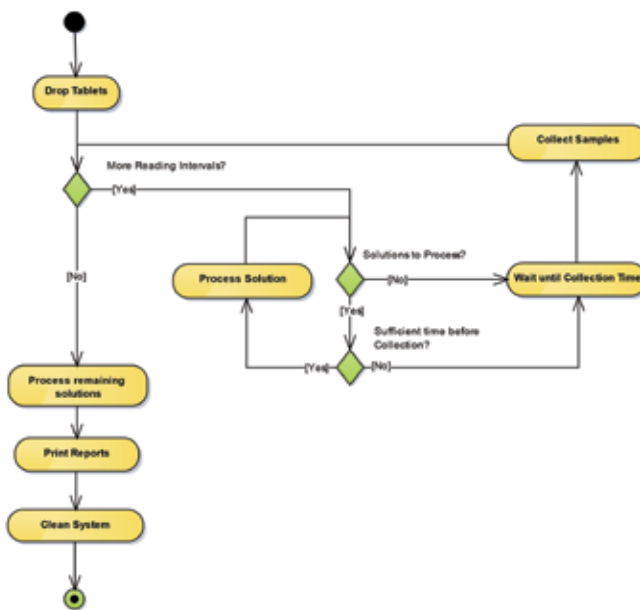


Figure 3. Flowchart of automated sequence.

Diluted solutions (sample, standard, and blanks) are aspirated then dispensed in the appropriate order into a port on the ASP2000 connected to a flow cell in the spectrophotometer, and measurements are performed on the spectrophotometer. Dissolution results are calculated and displayed in real time. The complete system is controlled by the IDIS software, which also manages the calculations and data record storage using an Oracle database. The entire sequence of operations (collection, filtration, media replacement, dilution, measurement, and calculation of results) is automated and requires no operator intervention.

AUTOMATED DISSOLUTION TESTS FOR METFORMIN HYDROCHLORIDE

Commercial samples of metformin hydrochloride extended-release 500-mg tablets were analyzed in accordance with *USP* (6) using the automated system described.

The test used 1000 mL of pH 7.4 ± 0.05 phosphate buffer (6.8 ± 0.1 g of potassium dihydrogen phosphate and 1.58 ± 0.05 g of sodium hydroxide in 1000 mL of deionized degassed water). The system was configured with paddles (1, 2); 10 mL of sample was collected at each interval (six

samples) at 15 min, 30 min, and 1, 2, 3, 4, 6, 8, 10, 12, and 14 h. After each sampling, the volume removed from the vessel was automatically replaced by the system.

The graphically configured method is shown by Figure 4. A method was configured to collect samples at multiple time points; prepare standards, blanks, and samples by dilution; and inject each solution into the UV. After injection, UV measurement was performed; dissolution results were calculated and displayed in real time. The complete process is streamlined and automated; this was not previously achievable.

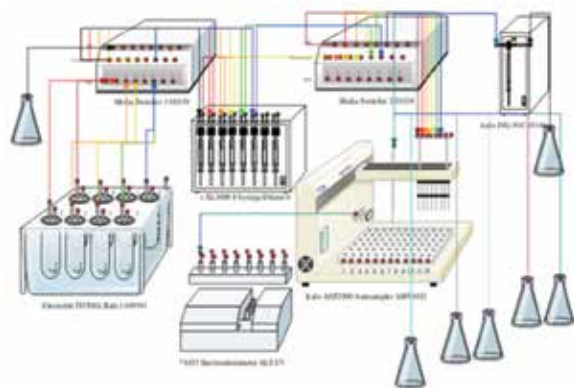


Figure 4. Method configuration schematic. Actual representation of the method configuration for the analysis.

Samples collected were diluted 1 in 10, then further diluted 1 in 10 to achieve a dilution of 1 in 100, shown by schematic Figure 2. Stock standards were prepared by accurately weighing and dissolving 10 mg of metformin hydrochloride in 100 mL of DI water. Stock solutions were further diluted by the system (1 in 20), and the diluted standards and samples were injected and measured on the spectrophotometer at 233 nm. Prior to reading standards and samples, blank measurements were performed. Dissolution results were calculated as the experiment proceeded, and the profile was displayed in real time.

Results and dissolution profiles were calculated and plotted in real time, and reports were generated automatically at the end of the analysis. The UV absorbance readings for Salicylic Acid are shown in Tables 1 and 2, and Table 3 shows the percent dissolved results for metformin hydrochloride.

System Control and Synchronization of Collection with Sample Processing

Full automation of dissolution has very complex procedures. The time points of sampling, dilution, and injection must be controlled precisely. If this is not coordinated, there will be timing conflicts between the processing performed in real time and collection, which must be precise according to pharmacopeia.

The IDIS software controls the instruments and uses an innovative algorithm to coordinate and optimize scheduling to improve efficiency and avoid conflicts between the

collection and the sample dilution and measurement times (Figure 3). Without the scheduler (deferred data collection), processes would have to be performed within large time intervals, which limits sampling at short time intervals.

Compliance with Pharmacopeia Guidelines

Strict quality control is required for automation of dissolution in NIFDC. The automated system complies with FDA 21 CFR guidelines, allowing use in pharmacopeia-compliant laboratories, importantly quality control environments. The system provides four-level right access in the system: Administration, Development, Application, and Instrumentation with electronic signing.

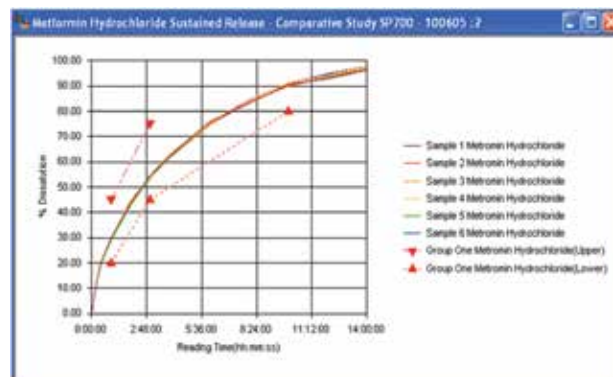


Figure 5. Percent dissolution graph from the automated system for metformin hydrochloride dissolution profile with high/low specification compliance error limits in red.

VALIDATION

The automated system was validated in accordance with the United States, European, and Chinese pharmacopeias (1–3) using 300-mg salicylic acid tablets, LOT 100103-200610 (NIFDC, China), dissolved in 900 mL of pH 6.8 phosphate buffer medium prepared with 6.8 ± 0.1 g of potassium dihydrogen phosphate and 1.58 ± 0.05 g of sodium hydroxide (both from Sinopharm Chemical Reagent Co., Ltd, China) in 1000 mL of deionized, degassed water (3). The system was configured with a bath paddle speed of 100 rpm (1, 2) at a temperature of 37 ± 0.5 °C. A 10-mL sample was collected at each time point, diluted 1 in 5, and mixed automatically. Two individually prepared standards (15 mg in 200 mL) were diluted manually 1 in 5. The diluted standards and samples were injected by the system into the UV spectrophotometer, measured at 296 nm, and dissolution results were calculated.

A 1% standard check was configured to verify the standard measurement. For comparison, samples were taken manually from the bath vessels at exactly the same time they were collected by the system. These samples were diluted and measured manually and compared to results produced by the automated system.

Readings obtained manually by measuring in a 0.3-cm flow cell (Shimadzu Corporation, Kyoto Japan) and from the automated system after preparation (diluted 1 in 5 and mixed) are shown in Table 1.

Table 1. Results for Two Individually Prepared Standards Measured Manually and with the Automated System after Dilution

Standard Samples		Manual Sampling		Automated Dissolution System	
		Absorbance (0.3-cm cell)	Response	Absorbance (diluted 5-fold, 1-cm cell)	Response Difference (%)
Control 1	15.05	0.594	0.99	0.393	0.99
Control 2	15.06	0.599		0.397	

Table 2. Manual and Automated Results for Salicylic Acid Dissolution Analysis

Vessel No.	Manual Sampling		Automated Dissolution System		% Deviation
	Absorbance (0.3-cm cell)	% Dissolution	Absorbance (diluted 5-fold, 1-cm cell)	% Dissolution	
1	0.729	28	0.475	27	1.6
2	0.778	30	0.505	29	2.0
3	0.759	29	0.495	28	1.5
4	0.724	27	0.471	27	1.8
5	0.722	27	0.466	27	2.5
6	0.692	26	0.451	26	1.6

Table 3. Results for Commercially Available Metformin Hydrochloride 500-mg Extended-Release Tablets from Three Different Manufacturers

Sampling Time	Manufacturer					
	A		B		C	
	Manual	ASP2000 System	Manual	ASP2000 System	Manual	ASP2000 System
15 min	—	14%	—	12%	—	13%
30 min	—	21%	—	20%	—	23%
1 h	31%	31%	30%	30%	33%	34%
2 h	—	46%	—	44%	—	48%
3 h	57%	56%	54%	54%	58%	59%
4 h	—	65%	—	62%	—	67%
6 h	—	78%	—	75%	—	79%
8 h	—	87%	—	84%	—	87%
10 h	93%	93%	90%	91%	91%	90%
12 h	—	97%	—	94%	—	92%
14 h	—	100%	—	97%	—	92%

CONCLUSION

The results of the validation show very good agreement with manual procedures performed using the same stock standard solution. Acceptable repeatability was obtained from multiple measurements. Repeated tests involved loading created standard methods and following the on-screen instructions, which makes the system very robust for use in a QC environment. Reports were professionally presented in different graphical formats, as shown by a typical report graph in Figure 5. In practice, trained analysts should achieve the same results thus eliminating user variability.

Repeatability of results shows low CV, even with dilution ratios of 1 in 100. The results for all samples at 1, 3, and 10 h agree within 1% of the manual method. With the automated system, it was possible to measure more time-point readings due to a high degree of automation and no analyst intervention, hence the number of sampling times was increased (Table 3). The objective to develop an automated method that could collect and dilute samples and measure automatically was demonstrated and validated.

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