

Development of In Vivo Predictive pH-Gradient Biphasic Dissolution Test for Weakly Basic Drugs: Optimization by Orthogonal Design

Xiaowei Fan¹, Shengying Shi¹, Junlin He¹, Jia Deng^{2,3*}, Jingou Ji^{1*}

¹College of Chemistry and Chemical Engineering, Chongqing University, Chongqing, China.

²Chongqing Key Laboratory of Natural Products, College of Environment and Resources, Chongqing Technology and Business University, Chongqing, China.

³Central Nervous System Drug Key Laboratory of Sichuan Province, Luzhou, Sichuan, China.

e-mail: 725_tiger@sina.com;
jiadeng2011@hotmail.com

ABSTRACT

The aim of this study was to develop a method for set up and optimization of a pH-gradient biphasic dissolution model by orthogonal test design in light of the correlation with published in vivo data of ketoconazole (KTZ). A pH-gradient biphasic dissolution test was designed with a sequential pH-gradient in the aqueous phase to simulate stomach, duodenum, jejunum, and ileum, and the organic phase was added in simulated small intestine conditions. The model was optimized by orthogonal test design with three factors and three levels and correlating with the published pharmacokinetic data of pure drug. The optimized dissolution conditions were 30 rpm, 100 mL of an organic volume, and pH 5.5, 6.5, and 6.8 in the pH-gradient aqueous phase in USP apparatus 2. Under these conditions, KTZ dissolution displayed a good linear relationship with in vivo absorption ($R^2 = 0.85$). This study indicates that this methodology is feasible to develop an in vivo predictive dissolution test.

KEYWORDS: pH-gradient biphasic dissolution test, in vitro-in vivo relationship (IVIVR), weakly basic drug, predictive dissolution test, ketoconazole, dissolution

INTRODUCTION

Oral absorption of drugs is an extremely complicated process that is influenced by physiochemical properties of the drug formulation and physiological conditions of the gastrointestinal (GI) tract (1). Edwards in 1951 first proposed that drug dissolution would play a critical role in oral absorption, and this relationship has been of great interest in the past decades (2, 3). In vitro dissolution testing provides considerable information for in vivo drug performance by establishing an in vitro-in vivo relationship (IVIVR). Noticeably, when dissolution tests have inaccurate discrimination for the performance of candidate formulations between in vitro and in vivo studies, the predictive results may be completely misleading. Thus, it is desirable that changes in in vivo drug dissolution are reflected by the corresponding in vitro drug release test.

The oral absorption of Biopharmaceutics Classification System (BCS) class II drugs is mainly limited by solubility and dissolution, which may be required to establish the IVIVR (4, 5). In particular, weakly basic drugs are

significantly influenced by physiological conditions in the GI tract, especially for dynamic pH conditions. Weak bases freely dissolve at gastric pH, but not at intestinal pH due to a supersaturation-precipitation process that greatly affects drug bioavailability. Some dissolution tests may employ simple and single-pH non-physiologic buffers that do not accurately reflect in vivo dynamic situations (6, 7). To bridge the gap between in vitro and in vivo conditions, many attempts have been made to replicate the GI process of weak bases by developing various biorelevant dissolution methods (8, 9). Owing to the lack of drug removal from the system and challenges associated with simulated absorption across the intestinal membrane, in vitro dissolution may overestimate the in vivo situation.

A biphasic dissolution test offers the advantage of maintaining sink conditions and having an absorptive phase, which has had increasing attention (10). A biphasic dissolution test can simulate drug dissolution and absorption in the GI tract by the implementation of an immiscible organic phase acting as an absorptive sink over the aqueous solution. Moreover, the setup and handling of biphasic tests are relatively simple and cost

*Corresponding authors

effective, and drug release and partitioning are detected simultaneously within a single vessel (11). Previous studies have reported the development of various biphasic dissolution models and their correlation to pharmacokinetic (PK) data for different dosage forms (12–17). A biphasic system with a single pH of aqueous phase was developed by combining United States Pharmacopeia (USP) apparatus 2 and 4 methods to discriminate three celecoxib formulations and obtain a consistent ranking between drug concentrations of organic phase and in vivo PK parameters (12). To better mimic drug behaviors in the GI tract, a pH-adjusted biphasic dissolution system was developed to differentiate between four modified release formulations prepared with two weakly basic drugs (dipyridamole and BIMT 17) (13). The pH-adjusted biphasic test provided a ranking prediction with respect to in vivo absorption, but not in dissolution tests with different media at a constant pH. However, there are still some questions to be considered and solved. Firstly, there is a lack of studies that develop a method for setup and optimization of a biphasic test because the experimental parameters may have a critical influence on drug performance (11, 18). Secondly, pH-adjusted biphasic dissolution tests involve the addition of organic phase at gastric pH, which might result in an overestimated prediction of in vivo performance due to supersaturation of weak bases, which could lead to overestimation of oral bioavailability if the organic phase is added prematurely.

Based on the aforementioned concerns, ketoconazole (KTZ), a BCS II weakly basic drug with broad-spectrum antifungal activity, was selected as a model drug. The aim of the present study was to develop a method to set up and optimize a pH-gradient biphasic dissolution model by orthogonal test design in light of the correlation with published in vivo data of KTZ.

MATERIALS AND METHODS

Materials

KTZ was purchased from Wuhan Dahua Weiye Medicine Chemical Co., Ltd. (Wuhan, China). 1-octanol, hydrochloric acid (HCl), sodium dihydrogen phosphate dihydrate, sodium hydroxide (NaOH), and sodium chloride were obtained from Sichuan Kelun Pharmaceutical Co., Ltd. (Chengdu, China). Hard gelatin capsules (size 0) were donated by Suzhou Capsugel Ltd. (Suzhou, China). All other reagents used were of analytical grade.

Solubility Determination

Excessive KTZ was added to 10 mL of five different dissolution media (gastric buffer pH 2.0, phosphate buffer pH 5.5, 6.5, and 1-octanol). The solubility test was performed at 37 °C and 160 rpm for 24 h using

a horizontal shaker (THZ-98AB, Shanghai Yiheng Scientific Instrument Co. Ltd, Shanghai, China) ($n = 3$). Saturated solutions were filtered through a 0.45- μm membrane filter. Drug concentration was measured by ultraviolet (UV) spectrophotometry at 224 nm (T6, Beijing Puxi General Instrument Co. Ltd, Beijing, China).

pH-Gradient Biphasic Dissolution Test

Considering the physiological pH-change in the GI tract which significantly influences KTZ dissolution, a pH-gradient biphasic dissolution test was proposed (Fig. 1). The test was performed in USP apparatus 2 (RCZ-6B3, Shanghai Huanghai Pharmaceutical Inspection Instrument Co. Ltd, Shanghai, China) combining with a pH controlling device (PHS-2F, Shanghai INESA Scientific Instrument Co., Ltd, Shanghai, China).

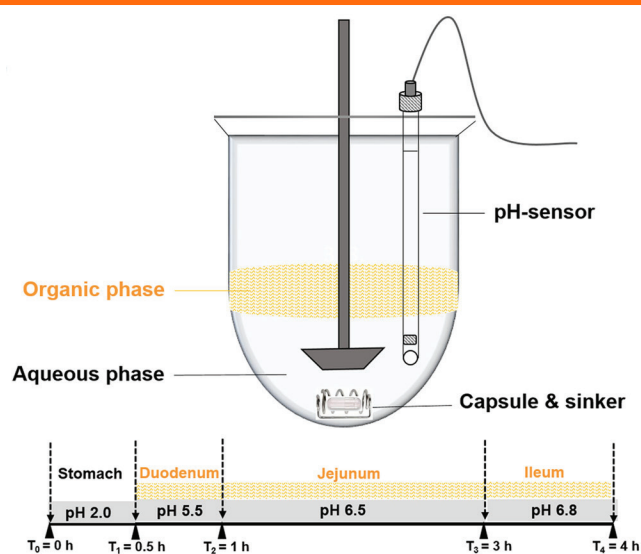


Figure 1. Schematic diagram of a pH-gradient biphasic dissolution system.

The gastric buffer was prepared with 250 mL of 0.01 N HCl with 0.1 M NaCl (pH 2.0) and used to simulate drug dissolution in the stomach for 30 min (19). The pH of the aqueous medium was adjusted to duodenal pH by using 5 M NaOH with a fresh sampling needle, and 1-octanol as the upper organic phase was added to simulate the intestinal membrane as an absorptive sink. 1-octanol was saturated with water prior to use to keep a constant volume during the experiment. Subsequently, the pH of the aqueous phase was readjusted to 6.5 to mimic the jejunum for 2 h, and then the final pH increased to 6.8 for 1 h. The temperature was maintained at 37 °C.

For the dissolution test, 100 mg of pure KTZ in a gelatin capsule was used with a sinker ($n = 3$); 3-mL samples were withdrawn manually from the aqueous phases at 5, 10, 20, 30, 40, 50, 60, 70, 80, 90, 105, 120, 150, 180, 210, and 240 min, while 2-mL samples were collected

in the organic phase at 40, 50, 60, 70, 80, 90, 105, 120, 150, 180, 210, and 240 min. Each sample was replaced with the same volume of fresh media. Samples collected from the aqueous phase were passed through a 0.45- μ m membrane filter, and samples from the organic phase were centrifuged at 12,000 rpm for 20 min (TG-16, Gongyi Yuhua Instrument Co. Ltd, Gongyi, China). The drug concentration was determined by UV assaying at 224 nm.

Optimization Strategy

To obtain an optimal dissolution model for good prediction of in vivo drug performance, optimization was carried out using a simple and efficient orthogonal experimental design. Rotating speed of the paddle (factor A), organic volume (factor B), and pH gradients of the aqueous phase (factor C) are most likely to influence the dissolution process, so these were selected as three factors. The orthogonal experiment with three factors and three levels, as shown in Table 1, was first used for optimization of the biphasic dissolution test. Nine trials were performed based on the $L_9(3^4)$ matrix with a minimal number of trials by testing combinations (20). Correlation coefficient (R^2) of a linear relationship between in vitro drug dissolution and the reported in vivo KTZ data obtained from (21) was selected as the evaluation index. The closer to 1 the R^2 value was, the better the predictive efficiency.

Table 1. Three Factors (A, B, and C) and Three Levels of the Orthogonal Test Design

Level	A (Rotating Speed, rpm)	B (Organic Volume, mL)	C (pH-Gradient Aqueous Phase)
1	30	75	5.5, 6.5, 6.8
2	40	100	6.0, 6.5, 6.8
3	50	125	6.5, 6.5, 6.8

Data Analysis

All data were expressed as mean \pm standard deviation (SD). Linear regression analysis was used to evaluate the relationship between the percentage of drug release and the percentage of drug absorbed (F_a) at corresponding time points. F_a was calculated via Microsoft Excel 2013 (Redmond, WA, USA) based on the reported in vivo data from female rats using the Wagner-Nelson method (21, 22):

$$F_a = \left[\frac{C_{(t)} + k_e \text{AUC}_{(0-t)}}{k_e \text{AUC}_{(0-\infty)}} \right] \times 100$$

In this equation, F_a is the fraction of drug absorbed; $C_{(t)}$ is the drug concentration at time point t ; k_e is the elimination rate constant; AUC_{0-t} is the area under the plasma concentration-time curve from zero to time t ; and $\text{AUC}_{0-\infty}$ is the area under the curve from zero to infinity.

RESULTS AND DISCUSSION

Solubility

KTZ showed highly pH-dependent solubility in the physiologically relevant pH range (Table 2). As a weakly basic drug, KTZ had the highest solubility in the acidic media, with 8.2 mg/mL dissolved in pH 2.0, which was used to mimic pH conditions of a healthy fasted stomach. In contrast, KTZ showed a dramatic decrease in solubility to 9.5 μ g/mL in pH 5.5 buffer, following a slow fall from 3.9 to 3.5 μ g/mL between pH 6.5 and pH 6.8. KTZ had high solubility of 5.6 mg/mL in 1-octanol due to its hydrophobic structure (logP 3.9) (23).

Table 2. Solubility of Ketoconazole in Dissolution Media at 37 °C

Media	Solubility, mean \pm SD ($n = 3$)
1-Octanol	5.6 \pm 0.5 mg/mL
Gastric buffer pH 2.0	8.2 \pm 0.2 mg/mL
PBS pH 5.5	9.5 \pm 0.5 μ g/mL
PBS pH 6.5	3.9 \pm 0.6 μ g/mL
PBS pH 6.8	3.5 \pm 0.6 μ g/mL

PBS: phosphate buffer solution.

Optimization Study

The orthogonal experimental design was used to select the optimum dissolution conditions. The results of nine groups of experiments under different conditions are shown in Table 3. The k (k_1 , k_2 , or k_3) value was the mean of the sum of the three R^2 values for levels 1–3 of each factor, which revealed the change of predictive efficiency. The effects of levels 1–3 on drug dissolution were factor A (rotating speed of paddle): 1 > 2 > 3; factor B (organic volume): 2 > 1 > 3; factor C (pH gradients of aqueous phase): 1 > 3 > 2. The range (R) was the difference between the maximum and minimum value of k_1 , k_2 , and k_3 in the same table column, which reflected the impact degree of different factors. The larger the R value of one factor, the greater the impact on drug dissolution and in vivo prediction. According to the R value in Table 3, the order of three factors influencing drug dissolution and in vivo prediction was C > B > A. Therefore, the optimized dissolution parameters were $A_1B_2C_1$; namely, rotating speed is 30 rpm, the organic volume is 100 mL, and the pH-gradient aqueous phase is pH 5.5, 6.5, and 6.8.

pH-Gradient Biphasic Dissolution Test

To validate the prediction potential of the optimized dissolution test, KTZ dissolution was performed in the optimum pH-gradient biphasic dissolution test in triplicate. The dissolution profiles of pure drug are presented in Figure 2. Drug dissolution reached 100%

Table 3. Experimental Design and Results of Orthogonal Test Design $L_9(3^4)$

Test No.	A (Rotating Speed, rpm)	B (Organic Volume, mL)	C (pH-gradient aqueous phase)	D Blank	R^2
1	30	75	5.5, 6.5, 6.8	1	0.81
2	30	100	6.0, 6.5, 6.8	2	0.65
3	30	125	6.5, 6.5, 6.8	3	0.63
4	40	75	6.0, 6.5, 6.8	3	0.66
5	40	100	6.5, 6.5, 6.8	1	0.73
6	40	125	5.5, 6.5, 6.8	2	0.65
7	50	75	6.5, 6.5, 6.8	2	0.64
8	50	100	5.5, 6.5, 6.8	3	0.83
9	50	125	6.0, 6.5, 6.8	1	0.44
k_1	0.70	0.70	0.76		
k_2	0.68	0.74	0.58		
k_3	2.64	0.57	0.67		
R	0.06	0.16	0.18		
OC	A ₁	B ₂	C ₁		

Note: k_1 , k_2 , and k_3 are mean values of the sum of level 1–3 for each factor (A–C); R is the difference between maximum and minimum value of k_1 , k_2 , and k_3 in the same column; R^2 : correlation coefficient; OC: optimized condition. oxide.

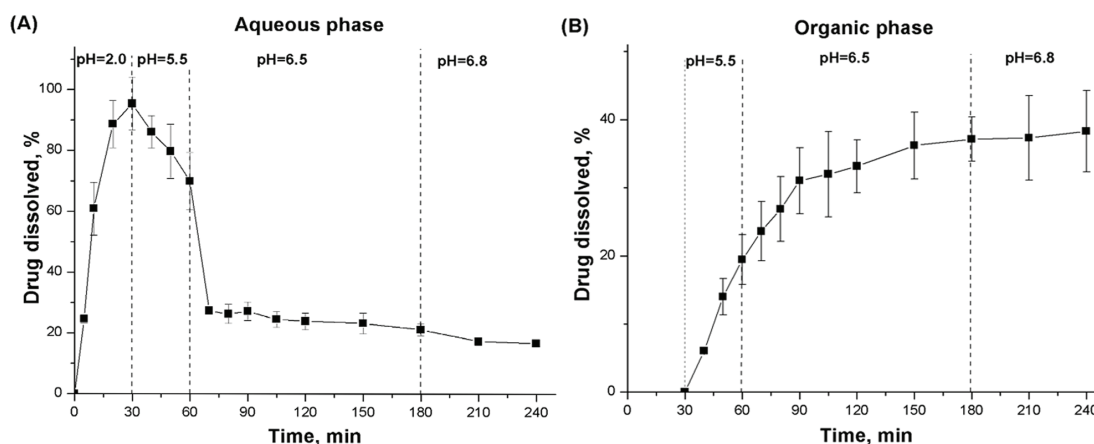


Figure 2. Dissolution profiles of pure ketoconazole in the optimized pH-gradient biphasic dissolution test in the aqueous phase (A) and organic phase (B) (gastric buffer pH 2.0 for 30 min followed by pH adjustment to 5.5 ± 0.05 for 30 min, 6.5 ± 0.05 for 2 h, and 6.8 ± 0.05 for 1 h, respectively). Dashed vertical lines represents the pH changes.

in the gastric buffer pH 2.0 due to high solubility in the acidic medium. After pH adjustment, drug concentration dramatically declined with increasing pH from 5.5 to 6.8 in the aqueous phase, even to below 20% in pH 6.8 medium (Fig. 2A). This could be attributed to rapid drug precipitation caused by low solubility of KTZ in the high pH media. In contrast, the amount of drug released gradually increased in the organic phase (Fig. 2B). It is well known that the small intestine is a main absorption site for most drugs (24). The addition of the organic phase after pH change was to simulate the intestinal absorption by removing dissolved drug from the aqueous phase and into the organic phase. Moreover, a good linear

relationship was found between in vitro dissolution and in vivo F_a ($R^2 = 0.85$) (Fig. 3), which was higher than the R^2 values obtained for all other conditions shown in Table 3.

CONCLUSION

An orthogonal design using experimental data combined with reported in vivo data was used to set up and optimize a pH-gradient biphasic dissolution test. The optimized experimental parameters were obtained based on R^2 of a linear relationship between in vitro and in vivo dissolution of KTZ. The highest R^2 was found when KTZ was tested in the optimized pH-gradient biphasic dissolution test, indicating feasibility of the method. In the next study,

different KTZ formulations will be used to further identify the discriminative and predictive ability of the proposed biphasic dissolution test.

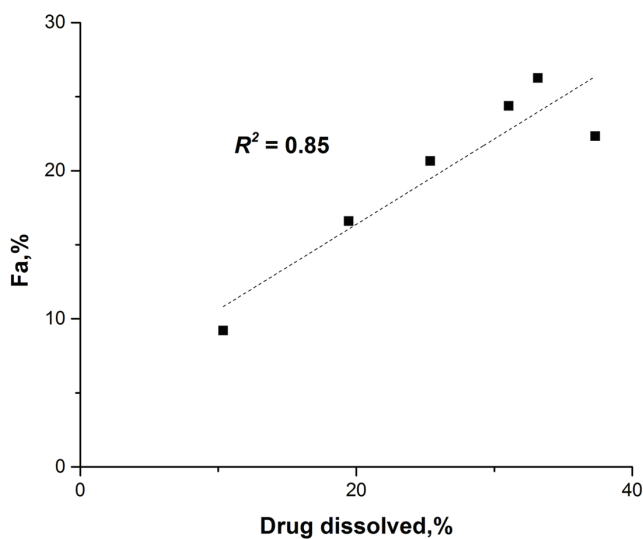


Figure 3. Relationship of in vitro dissolution of ketoconazole in the organic phase of optimized pH-gradient biphasic test plotted against in vivo absorption (Fa) calculated according to data reported in female rats (21).

FUNDING

This work was financially supported for purchasing materials and assay determination by the Science and Technology Research Program of Chongqing Municipal Education Commission (Grant No. KJQN201800820); the Chongqing Research Program of Basic Research and Frontier Technology (Grant No. cstc2019jcyj-msxmX0753); the Scientific Research Project of Chongqing Technology and Business University (Grant No. 1951066); the open fund of the Key Laboratory of Natural Medicine Research of Chongqing Education Commission (Grant No. KFJJ2019093); the Open Project of Central Nervous System Drug Key Laboratory of Sichuan Province (Grant No. 200024-01SZ); and the Open Project of Central Nervous System Drug Key Laboratory of Sichuan Province (Grant No. 200024-01SZ).

CONFLICT OF INTERESTS

The authors disclosed no conflicts of interest related to this article.

REFERENCES

- Jambhekar, S. S.; Breen, P. J. Drug dissolution: significance of physicochemical properties and physiological conditions. *Drug Discov. Today*. **2013**, *18*, 1173–1184. DOI: 10.1016/j.drudis.2013.08.013.
- Edwards, L. J. The dissolution and diffusion of aspirin in aqueous media. *Trans. Faraday Soc.* **1951**, *47*, 1191–1210. DOI: 10.1039/tf9514701191.
- Dokoumetzidis, A.; Macheras, P. A century of dissolution research: from Noyes and Whitney to the biopharmaceutics classification system. *Int. J. Pharm.* **2006**, *321*, 1–11. DOI: 10.1016/j.ijpharm.2006.07.011.
- Amidon, G. L.; Lennernäs, H.; Shah, V. P.; Crison, J. R. A theoretical basis for a biopharmaceutic drug classification: the correlation of in vitro drug product dissolution and in vivo bioavailability. *Pharm. Res.* **1995**, *12*, 413–420. DOI: 10.1023/A:1016212804288.
- Butler, J.; Dressman, J. B. The developability classification system: Application of biopharmaceutics concepts to formulation development. *J. Pharm. Sci.* **2010**, *99*, 4940–4954. DOI: 10.1002/jps.22217.
- Mudie, D. M.; Amidon, G. L.; Amidon, G. E. Physiological parameters for oral delivery and in vitro testing. *Mol. Pharm.* **2010**, *7*, 1388–1405. DOI: 10.1021/mp100149j.
- Park, K. Absence of in vivo-in vitro correlation in per-oral drug delivery. *J. Control. Release* **2014**, *180*, 150. DOI: 10.1016/j.jconrel.2014.03.020.
- Kostewicz, E. S.; Wunderlich, M.; Brauns, U.; Becker, R.; Bock, T.; Dressman, J. B. Predicting the precipitation of poorly soluble weak bases upon entry in the small intestine. *J. Pharm. Pharmacol.* **2004**, *56*, 43–51. DOI: 10.1211/0022357022511.
- Butler, J.; Hens, B.; Vertzoni, M.; Brouwers, J.; Berben, P.; Dressman, J.; Andreas, C. J.; Schaefer, K. J.; Mann, J.; McAllister, M.; Jamei, M.; Kostewicz, E.; Kesisoglou, F.; Langguth, P.; Minekus, M.; Mullertz, A.; Schilderink, R.; Koziol, M.; Jedamzik, P.; Weitschies, W.; Reppas, C.; Augustijns, P. In vitro models for the prediction of in vivo performance of oral dosage forms: Recent progress from partnership through the IMI OrBiTo collaboration. *Eur. J. Pharm. Biopharm.* **2019**, *136*, 70–83. DOI: 10.1016/j.ejpb.2018.12.010.
- Pestieau, A.; Evrard, B. In vitro biphasic dissolution tests and their suitability for establishing in vitro-in vivo correlations: A historical review. *Eur. J. Pharm. Sci.* **2017**, *102*, 203–219. DOI: 10.1016/j.ejps.2017.03.019.
- Locher, K.; Borghardt, J. M.; Frank, K. J.; Kloft, C.; Wagner, K. G. Evolution of a mini-scale biphasic dissolution model: Impact of model parameters on partitioning of dissolved API and modelling of in vivo-relevant kinetics. *Eur. J. Pharm. Biopharm.* **2016**, *105*, 166–175. DOI: 10.1016/j.ejpb.2016.06.008.
- Shi, Y.; Gao, P.; Gong, Y.; Ping, H. Application of a biphasic test for characterization of in vitro drug release of immediate release formulations of celecoxib and its relevance to in vivo absorption. *Mol. Pharm.* **2010**, *7*, 1458–1465. DOI: 10.1021/mp100114a.
- Heigoldt, U.; Sommer, F.; Daniels, R.; Karl-Gerhard, W. Predicting in vivo absorption behavior of oral modified release dosage forms containing pH-dependent poorly soluble drugs using a novel pH-adjusted biphasic in vitro dissolution test. *Eur. J. Pharm. Biopharm.* **2010**, *76*, 105–111. DOI: 10.1016/j.ejpb.2010.05.006.
- Deng, J.; Staufienbiel, S.; Hao, S.; Wang, B.; Dashevskiy, A.; Bodmeier, R. Development of a discriminative biphasic in vitro dissolution test and correlation with in vivo pharmacokinetic

- studies for differently formulated racecadotril granules. *J. Control. Release.* **2017**, *255*, 202–209. DOI: 10.1016/j.jconrel.2017.04.034.
15. Pestieau, A.; Lebrun, S.; Cahay, B.; Brouwers, A.; Streel, B.; Cardot, J. M.; Evrard, B. Evaluation of different in vitro dissolution tests based on level A in vitro-in vivo correlations for fenofibrate self-emulsifying lipid-based formulations. *Eur. J. Pharm. Biopharm.* **2017**, *112*, 18–29. DOI: 10.1016/j.ejpb.2016.10.030.
 16. Deng, J.; Staufenbiel, S.; Bodmeier, R. Evaluation of a biphasic in vitro dissolution test for estimating the bioavailability of carbamazepine polymorphic forms. *Eur. J. Pharm. Sci.* **2017**, *105*, 64–70. DOI:10.1016/j.ejps.2017.05.013.
 17. Xu, H.; Shi, Y.; Vela, S.; Marroum, P.; Gao, P. Developing quantitative in vitro-in vivo correlation for fenofibrate immediate-release formulations with the biphasic dissolution-partition test Method. *J. Pharm. Sci.* **2018**, *107*, 476–487. DOI: 10.1016/j.xphs.2017.06.018.
 18. Vangani, S.; Li, X. L.; Zhou, P.; Del-Barrio, M. A.; Chiu, R.; Cauchon, N.; Gao, P.; Medina, C.; Jasti, B. Dissolution of poorly water-soluble drugs in biphasic media using USP 4 and fiber optic system. *Clin. Res. Regul. Aff.* **2009**, *26*, 8–19. DOI: 10.1080/10601330902905887.
 19. Klein, S. The use of biorelevant dissolution media to forecast the in vivo performance of a drug. *AAPS. J.* **2010**, *12*, 397–406. DOI: 10.1208/s12248-010-9203-3.
 20. Fan, W.; Zhu, W.; Zhang, X.; Di, L. The preparation of curcumin sustained-release solid dispersion by hot melt extrusion-I. Optimization of the formulation. *J. Pharm. Sci.* **2020**, *109*, 1242–1252. DOI: 10.1016/j.xphs.2019.11.019.
 21. Zhou, Y.; He, P.; Liu, A.; Zhang, L.; Liu, Y. D.; Dai, R. Drug–drug interactions between ketoconazole and berberine in rats: pharmacokinetic effects benefit pharmacodynamic synergism. *Phytother. Res.* **2012**, *26*, 772–777. DOI: 10.1002/ptr.3621.
 22. Takka, S.; Sakr, A.; Goldberg, A. Development and validation of an in vitro-in vivo correlation for buspirone hydrochloride extended release tablets. *J. Control. Release.* **2003**, *88*, 147–157. DOI:10.1016/S0168-3659(02)00490-X.
 23. Ghasemi, J.; Saaidpour, S. Quantitative structure-property relationship study of n-octanol-water partition coefficients of some of diverse drugs using multiple linear regression. *Anal. Chim. Acta.* **2007**, *604*, 99–106. DOI: 10.1016/j.aca.2007.10.004.
 24. Murakami, T. Absorption sites of orally administered drugs in the small intestine. *Expert. Opin. Drug Discov.* **2017**, *12*, 1219–1232. DOI: 10.1080/17460441.2017.1378176.